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(54) Title: **CLONING, SEQUENCING AND EXPRESSION OF A COMAMONAS CYCLOPENTANONE 1,2-MONOOXYGENASE-ENCODING GENE IN ESCHERICHIA COLI**

(57) Abstract: Cyclopentanone 1,2-monooxygenase (CPMO) from *Comamonas* (previously *Pseudomonas*) sp. strain NCIMB 9872 carries out the second step of a degradation pathway that allows the bacterium to use cyclopentanol as a sole carbon source for growth. In the present invention there is reported the localization of the CPMO-encoding gene (*cpnB*) on a 4.3-kb *SphI* fragment, the determination of its sequence. The 550-amino acid CPMO polypeptide (*M_r* 62,111) encoded by the gene was found to have 36.5% identity with the sequence of cyclohexanone 1,2-monooxygenase (CHMO) of *Acinetobacter* sp. strain NCIMB 9871. The 62-kDa CPMO was expressed in *E. coli* as an IPTG-inducible protein.

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CLONING, SEQUENCING AND EXPRESSION OF A COMAMONAS
CYCLOPENTANONE 1,2-MONOOXYGENASE-ENCODING GENE IN

ESCHERICHIA COLI

TECHNICAL FIELD

5 The present invention relates to an isolated DNA encoding a cyclopentanone monooxygenase (CPMO), or an enzymatically active portion thereof, and expression vector and a transformed cell containing the isolates DNA.

BACKGROUND OF THE INVENTION

10 *Comamonas* (previously *Pseudomonas*) sp. NCIMB 9872 was one of the few microorganisms that have been characterized to produce a Baeyer-Villiger monooxygenase (BVMO; Griffin, M., et al., *Biochem. J.* **129**:595-603, 1972; Griffin, M., et al., *Eur. J. Biochem.* **63**:199-209, 1976; and Willetts, A., *Trends in Biotech.* **15**:55-62, 1997; for a recent review). BVMOs are flavoproteins that
15 mimic the classical Baeyer-Villiger organic chemical reaction which is a peracid-catalyzed oxidation of a ketone to an ester or lactone. The use of enzyme substitutes for the production of lactones in high yield and optical purity is an attractive feature in current trends of research and development toward replacing chemical methods with biological alternatives (Stinson, S.C., *Chem.*
20 *Eng. News*, 83-104, 1998). To date, the best characterized BVMO enzyme is that of cyclohexanone monooxygenase (CHMO) produced by *Acinetobacter* sp. NCIMB 9871 (Stewart, J.D., *Curr. Org. Chem.* **2**:195-216, 1998; Willetts, A., *Trends in Biotech.* **15**:55-62, 1997). This is also the only BVMO whose gene has been cloned and sequenced (Chen, et al., *J. Bacteriol.* **170**:781-789, 1988).
25 Recently, this valuable resource was used to engineer a "designer yeast" in a whole-cell approach to effect a variety of asymmetric Baeyer-Villiger oxidations (Stewart, J.D., et al., *J. Am. Chem. Soc.* **120**:3541-3548, 1998).

It would be highly desirable to be provided with a new CPMO having an increased enzymatic activity for growing cells in a medium containing
30 cyclopentanol or cyclopentanone as sole carbon source.

SUMMARY OF THE INVENTION

One aim of the present invention is to provide a new CPMO having an increased enzymatic activity for growing cells in a medium containing cyclopentanol or cyclopentanone as sole carbon source.

5 In accordance with the present invention there is provided an isolated DNA encoding a cyclopentanone monooxygenase (CPMO), or an enzymatically active portion thereof, the isolated DNA being characterized by the ability to hybridize specifically with the complement of the DNA represented in SEQ ID NO:8 under stringent hybridization conditions.

10 Also in accordance with the present invention, there is provided an isolated DNA, wherein it codes for a cyclopentanone monooxygenase (CPMO), and contains:

- (1) the nucleic acid sequence of SEQ ID NO:8;
- (2) a sequence corresponding to said nucleic acid sequence in
15 the scope of the degeneration of the genetic code; or
- (3) a sequence hybridizing under stringent conditions with the sequence from (1) or (2), and still coding for cyclopentanone monooxygenase (CPMO).

20 Still in accordance with the present invention, there is provided an isolated DNA encoding a cyclopentanone monooxygenase (CPMO), or an enzymatically active portion thereof, said isolated DNA having SEQ ID NO:8.

The present invention further provides an isolated DNA expression vector encoding an enzymatically active cyclopentanone monooxygenase (CPMO)
25 comprising a DNA characterized by a sequence as set forth in SEQ ID NO:8, or a portion thereof, said portion encoding said CPMO, in expressible form.

In accordance with the present invention, there is also provided a recombinant vector comprising the isolated DNA as described above, wherein the isolated DNA encodes cyclopentanone monooxygenase.

In a preferred embodiment of the present invention, the isolated DNA has a nucleic acid sequence of SEQ ID NO:8 or which, due to the degeneracy of the genetic code, is a functional equivalent thereof.

Also in accordance with the present invention, there is provided a
5 recombinant vector containing one or more copies of a recombinant DNA described above.

The recombinant vector may be a prokaryotic vector. The recombinant vector may also be a plasmid.

Therefore, in accordance with the present invention, there is also
10 provided a biologically functional plasmid or viral DNA vector, which contains a DNA as described above.

The present invention also provide a host cell comprising a recombinant vector as described above.

Accordingly, there is also provided a cell transformed with a heterologous
15 DNA expression construct encoding an enzymatically active cyclopentanone monooxygenase (CPMO) comprising a DNA characterized by a sequence as set forth in SEQ ID NO:8, or a portion thereof, said portion encoding said CPMO, in expressible form.

The cell may be a prokaryotic cell or it may be *E. coli*.

20 Still in accordance with the present invention, there is also provided a purified cyclopentanone monooxygenase (CPMO) having :

- a) an amino acid sequence as set forth in SEQ ID NO:5;
- b) an amino acid sequence encoded by a nucleic acid sequence as set forth in SEQ ID NO:8; or
- 25 c) an amino acid sequence encoded by a nucleic acid sequence hybridizing to a nucleic acid sequence complementary to the nucleic acid sequence of step b) above under stringent conditions, said amino acid sequence encoded in step c) having a same activity as the amino acid sequence in a).

The present invention also provides a recombinant cyclopentanone monooxygenase (CPMO) having an enzymatic activity superior to the one from a native *Pseudomonas*, and more preferably twice superior.

5 The recombinant cyclopentanone monooxygenase (CPMO) may be prepared from *Comamonas* sp. NCIMB 9872. The recombinant cyclopentanone monooxygenase (CPMO) has preferably a sequence as set forth in SEQ ID NO:5.

10 A method for growing cells *in vitro* in presence of cyclopentanol or cyclopentanone as sole source of carbon, said method comprising the steps of:

- a) transforming a cell with the expression construct described above; and
- b) growing the cell of step a) under suitable conditions in a medium containing cyclopentanol or cyclopentanone as a sole
15 source of carbon.

To increase this gene potential, according to the invention it is reported herein the cloning of a cyclopentanone monooxygenase (CPMO)-encoding gene (*cpnB*) from *Comamonas* (*Pseudomonas*) sp. NCIMB 9872, the determination of its DNA and surrounding sequence and expression of CPMO
20 activity and protein in *E. coli*.

BRIEF DESCRIPTION OF THE DRAWINGS

Fig. 1 illustrates the first two steps of cyclopentanol degradation by *Pseudomonas* sp. NCIMB 9872;

25 Fig. 2 illustrates the genetic organization in *Comamonas* sp. NCIMB 9872 in the *SphI* fragment containing cyclopentanone monooxygenase-encoding gene (*cpnB*) and additional open reading frames;

Fig. 3 illustrates an alignment of the amino acid sequence of the CPMO of *Comamonas* sp. NCIMB 9872 with that of CHMO from *Acinetobacter* sp.

NCIMB 9871 and a steroid monooxygenase (STMO) from *Rhodococcus rhodochromus*;

Fig. 4 illustrates a SDS-PAGE of crude extracts from *E. coli* (pCMP201); and

5 Fig. 5 illustrates CPMO-encoding gene, designated cpnB.

DETAILED DESCRIPTION OF THE PREFERRED EMBODIMENT

Cloning of the *Comamonas* sp. NCIMB 9872 CPMO-encoding gene

Pseudomonas sp. NCIMB 9872 (henceforth strain 9872) identified as a *Comamonas* by 16S rDNA sequencing in this study, was purchased from the
10 National Collections of Industrial and Marine Bacteria Ltd (NCIMB, Aberdeen, Scotland) and grown at 30°C in Luria-Bertani (LB) broth (Sambrook, J., et al., Molecular cloning: a laboratory manual, 2nd ed. Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N. Y, 1989), or mineral salt medium (MSM), pH 7.0, containing 2 ml of cyclopentanone. The MSM recipe contains per liter: 1.0 g of
15 NH₄NO₃, 1.5 g of KH₂PO₄, 1.5 g of Na₂HPO₄, 0.2 g MgSO₄·7H₂O, 0.01 g of CaCl₂·2H₂O, 0.005 of FeSO₄·7H₂O, 0.002 g of MnSO₄·4H₂O and 0.1 g of yeast extract. Agar was added to 1.5% for plates. Genomic DNA of strain 9872 was prepared by the Marmur method (Marmur, J., *J. Mol. Biol.* **3**: 208-218, 1961). At first, a Southern hybridization of DNA digested with *Bam*HI was carried out
20 using the *Acinetobacter* NCIMB 9871 CHMO-containing gene as probe. Since there was no positive result (hybridization conditions carried out at 65°C) the CPMO protein was purified in order to obtain an N-terminal amino acid sequence. The purification of CPMO protein from cyclopentanone-grown cells was according to Griffin and Turgill (Griffin, M., et al., *Eur. J. Biochem.* **63**:199-
25 209, 1976). Using an automated protein sequencer (Perkin-Elmer model 477) a 40-residue amino-terminal sequence of the purified CPMO was obtained (Fig. 2). This sequence, longer by 11 amino acids, is in perfect agreement with that reported previously from the same organism (Willets, A., *Trends in Biotech.* **15**:55-62, 1997). Two degenerate oligodeoxynucleotide primers (5'-
30 ACIACIATGA CIACNATGAC-3' (SEQ ID NO:1) and 5'-ARRTGRTAIA

RYTGRTA-3' (SEQ ID NO:2), corresponding to amino acids 2-8 and 35-40, respectively) were synthesized to amplify a 116-bp product from total DNA prepared from strain 9872. The PCR amplification was performed in a Perkin Elmer-Model 2400 Thermal Cycler™ and the amplification conditions were 94°C for 1 min, 50°C for 1 min and 72°C for 1 min for 30 cycles. The amplified product was cloned directly in the pXcmkn12 vector (Cha, J., et al., *Gene* **136**, 369-370, 1993), transformed in *E. coli* JM109 and the resulting plasmid was designated pCMP10. Before using the amplified product as a gene probe its nucleotide sequence was confirmed. Nucleotide sequencing was determined by the Taq DyeDeoxy terminator cycle sequencing kit and the ABI Prism 310 Genetic Analyzer (Perkin Elmer). Plasmid isolation was performed by the method of Birnboim and Doly (Birnboim, H. C., and J. Doly, *DNA. Nucleic Acids Res.* **7**:1513-1523, 1979).

In Fig. 2, Orf1 is most likely a transcriptional activator of the NtrC-type (Morett, E., L. Segovia, *J. Bacteriol.* **175**:6067-6074, 1993). The amino acid sequence of ORF1 (C-terminal 391-amino acids) showed 38-40% identity to equivalent regions of proteins such as NTRC_ECOLI (Nitrogen regulation protein NR(I) from *E. coli*; Miranda, et al., *The complete nucleotide sequence of the glnALG operon of Escherichia coli K12* **15**:2757-2770, 1987), ACOR_ALCEU (Acetoin catabolism regulatory protein from *Ralstonia eutropha*; Kruger, N., et al., *J. Bacteriol.* **179**:4391-4400, 1992). The amino acid sequence of ORF2, showing similarity to enzymes of the short-chain alcohol dehydrogenase family (Jornvall, H., et al., *Biochemistry* **34**: 6003-6013, 1995), is most homologous (45-46% identity) to a putative oxidoreductase CY39.16C of *Mycobacterium tuberculosis* (Swiss Prot sp:Q10855) and fadG3 of *M. tuberculosis* (GenBank accession number Z74025). For *Pseudomonas* sp. strain HI-201 the *lacZ*-Km^r cassette from pKOK6.1 (Kokotek, W., et al., *Gene* **84**: 467-471, 1989) was inserted into *cpnB* at the *Nsi*I site.

In Fig. 2, the following terms are defined as follows: t_{fd}, transcriptional termination sequence of phage fd; Km^r, kanamycin resistance gene, *lacZ*, gene encoding β-galactosidase. Genes and markers are indicated with arrows.

To clone the CPMO-containing gene, the DNA insert from pCMP10 was amplified, labeled by the digoxigenin-11-UTP system according to manufacturer's instructions (Boehringer Mannheim GmbH) and used to probe a Southern hybridization of strain 9872 genomic DNA digested with various restriction enzymes (*Bam*HI, *Eco*RI, *Hind*III, *Kpn*I, *Nhe*I, *Pst*I, *Sal*I, *Sph*I and *Xba*I). As a result, a single hybridizing band of ca 4.3-kb *Sph*I fragment was obtained. Conditions of hybridization were as before. Subsequently, a purified 4.0- to 4.5-kb size fraction of *Sph*I-cut total DNA separated on a 0.8 % agarose gel was ligated to *E. coli* plasmid pUC18, which had been linearized and dephosphorylated. A clone containing the 4.3-kb insert was screened by colony hybridization using the PCR product as a probe; this recombinant plasmid was designated pCMP200.

DNA sequence of the CPMO-encoding gene (*cpnB*) and the flanking region

Nucleotide sequencing of the CPMO-encoding gene was initiated by using a primer designed from the sequence of the PCR product cloned in pCMP10 and further extended using oligonucleotides derived from the new sequence. Both DNA strands of the *Sph*I fragment were sequenced and found to consist of 4281 base pairs (bp). The sequence was analyzed by GENETYX-Mac (Software Development Co., Ltd. Chiba, Japan) and the BLAST program (Altschul, S. F., et al., *Nucleic Acids Res.* **25**:3389-3402, 1997). As a result three open reading frames (ORFs) arranged in the same direction were predicted (Fig. 2). The nucleotide sequence of the 1650-bp ORF encoding CPMO is preceded by a partial ORF1 (1173-bp) coding for the C-terminus of an NtrC-type transcriptional activator (Miranda, et al., *The complete nucleotide sequence of the glnALG operon of Escherichia coli K12* **15**:2757-2770, 1987) and by a complete ORF2 (750-bp) coding for a homolog of the short-chain dehydrogenases/reductases (Jornvall, H., et al., *Biochemistry* **34**: 6003-6013, 1995). The two intergenic regions are 244-bp and 32-bp, respectively. The CPMO-encoding gene is referred to *cpnB* (cyclopentanone and *B* designates the second step of the degradation pathway, see Fig. 1) hereafter. In Fig. 5, the CPMO-encoding gene starts at nucleotide position 1822 and ends 3471 that does not include the stop codon. Accordingly, the boundary of *cpnA* is 3507-

4256. The partial open reading frame preceding *cpnB* is from 1-1174.

Fig. 1 has been adapted from Griffin, M., et al. (Griffin, M., et al., *Biochem. J.* **129**:595-603, 1972). The designated genes are: *cpnA* encoding cyclopentanol dehydrogenase; *cpnB* encoding cyclopentanone 1,2-
5 monooxygenase (CPMO). An alternative name for 5-valerolactone is 5-pentanolide. Subsequent reaction steps are the formation of 5-hydroxyvalerate, 5-oxovalerate, glutarate and finally acetyl CoA.

The amino acid sequence of the CPMO enzyme consists of 550 residues (Fig. 3). This sequence shows 36.5% identity and an additional 13.6% amino
10 acid similarity with the 543-residue CHMO of *Acinetobacter* sp. strain NCIMB 9871. An equally related protein (549 amino acids; 37.3% identity and 12.4% similarity) is the putative steroid monooxygenase (STMO) of *Rhodococcus rhodochrous* (Morii, S., et al., GenBank accession number AB010439, 1998). The latter enzyme carries out the oxidation of progesterone to produce
15 testosterone acetate. A CLUSTAL alignment of these three sequences gave 24.6% positional identity (Fig. 3).

In Fig. 3, asterisks indicate identical amino acids, dots indicated similar amino acids and dashes indicate gaps introduced to maximize the alignment. The amino-terminal peptide sequence confirmed by Edman degradation is
20 underlined. The locations of the consensus FAD fingerprint sequences as described by Eppink et al. (Eppink, et al., *Prot. Sci.* **6**:2454-2458, 1997) are as indicated. The conserved GD motif found in flavoprotein hydroxylases as a second FAD fingerprint is also indicated. Not shown is the DG motif of flavoprotein hydroxylases which has the sequence of chhhssDGxcSxhR. Lower
25 case letters identify certain residues types: h, hydrophobic residues, s, small residues, c, charged residues, and x, any residues. Note that a DG doublet is present in CPMO and STMO sequence.

A notable sequence motif present in CPMO and related proteins is the FAD-binding fingerprint (GXGXXG) that is similar to those found in flavoprotein
30 hydroxylases (Eppink, et al., *Prot. Sci.* **6**:2454-2458, 1997). Flavoprotein hydroxylases (eg. phenol hydroxylase, the structure is now known; Enroth, C.,

et al., *Structure* 6:605-617, 1998) are monooxygenases that catalyze the insertion of one atom of molecular oxygen into the substrate using NAD(P)H as electron donor. These proteins possess a conserved "Asp-Gly (DG)" motif for both FAD and NAD(P)H binding in between two fingerprint motifs for the FAD binding (fingerprint 1: GXGXXG; fingerprint 2: Gly-Asp [GD] motif). Sequence motifs in CPMO, STMO and CHMO differ from those in flavoprotein hydroxylases by having a repeated GXGXXG motif (amino acids 24 to 33 and 193 to 202 in CPMO numbering). The possibility that the second FAD fingerprint in CPMO and related proteins fulfils a dual role of FAD and NADPH binding awaits structural determination of a representative member of this family of proteins. It is reasonable to assume that a different mechanism in catalysis is reflected in the motifs seen in the two classes of proteins.

Expression of *cpnB* gene in *E. coli*

Two primers of the following sequence were synthesized to amplify the *cpnB* gene and the resultant 1.7-kb DNA fragment was cloned in the pSD80 plasmid to yield pCMP201. Plasmid pSD80 is a third generation derivative of the commercially available pKK223-3 vector (Pharmacia) that contains a *tac* promoter upstream of the multiple cloning site (MCS), an *unc* terminator sequence downstream of the MCS, and *lacI*^q elsewhere on the plasmid (Smith, S.P., et al., *Biochemistry* 35:8805-8814, 1996). The primers were: 5'-AAAAGGCCTG AACTTCAATT ATTTAGGAGA C-3' (SEQ ID NO:3) and 5'-AAAACTGCAG GAGTTGCACA ACAGAGTCTT AG-3' with built-in *Stu*I and *Pst*I restriction sites (underlined), respectively, to facilitate cloning at the compatible sites (*Sma*I and *Pst*I) of the pSD80 vector. Vent DNA polymerase (New England BioLabs, Beverly, MA) was used and the amplification conditions were 94°C for 1 min, 50°C for 1 min, and 72°C for 1 min, for 30 cycles. The amplified DNA fragment was purified from an agarose gel and digested with *Stu*I and *Pst*I. One of the resulting recombinant plasmids was designated pCMP201. By DNA sequencing it was established that no mutation had been introduced in the *cpnB* gene during PCR amplification.

Fig. 4 shows the production of a 60-kDa protein in a Coomassie blue-stained SDS-polyacrylamide gel of the crude protein extract prepared from *E. coli* JM109 (pCMP201) cells that were induced by 0.1 mM isopropyl-beta-D-thiogalactopyranoside (IPTG). The cells were induced at an absorbance (A₆₀₀) of 0.4 to 0.5 and the induction period was up to 4 hr. The observed molecular mass was in agreement with the predicted size of the 62-kDa CPMO. In the absence of IPTG, this protein band was not produced. Also, the CPMO enzyme activity was observed only in those cells grown in the presence of IPTG. CPMO activity was assayed at 25°C by measuring a decrease in absorbance at 340 nm in 50 mM phosphate buffer (pH 7.8) containing 1 µmol of cyclopentanone, 0.2 µmol of NADPH, and the crude enzyme extract prepared from *E. coli* JM109 (pCMP201). These cells were cultivated in 100 ml of LB medium containing 100 µg/ml of ampicillin at 25°C. The IPTG-induced cells were harvested by centrifugation, washed in 50 mM phosphate buffer (pH 7.2), resuspended in 1/20 volume of same buffer, and sonicated by four-20 sec bursts with a Braun-Sonifier™ 250 apparatus. After centrifugation for 30 min at 18,000 x g and at 4°C, the supernatant was used for determination of enzyme activity. One unit (U) of activity is defined as the amount of enzyme required to convert 1 µmol of substrate in 1 min. Protein concentration was determined by the method of Bradford (Bradford, M. M., *Anal. Biochem.* **72**: 248-254, 1976). As a result the specific activity of the CPMO enzyme was found to be 0.28 U/mg. The specific activity of CPMO in the native *Pseudomonas* was reported to be 0.11 U/mg (Griffin, M., et al., *Biochem. J.* **129**:595-603, 1972).

In Fig. 4, lane 1 has been loaded with extracts of IPTG-induced *E. coli* and lane 2 has been loaded with extracts of *E. coli* in absence of IPTG. M means molecular weight markers as indicated in kilo daltons. The arrow indicates the production of the desired 60-kDa protein.

Inactivation of *cpnB* gene

Pseudomonas sp. strain HI-201 was constructed by chromosomal inactivation of the *cpnB* gene using a *lacZ*-Km^r cassette from the mobilizable pKOK6.1 vector (Kokotek, W., et al., *Gene* **84**: 467-471, 1989). In pKOK6.1 the

lacZ gene is promoterless and in addition to Km^r it is ampicillin resistant (Ap^r). The *lacZ*-Km^r cassette was excised as a *Pst*I-fragment and inserted into the *Nsi*I site within the *cpnB* gene in pCMP200, yielding pCMP202. Electroporation of this plasmid into 9872 cells was carried out in the Gene Pulser™ (BioRads) and the parameters of electroporation were 2.5 kV, 25 uF and 200 ohm. The cells were initially washed with 1mM HEPES buffer and resuspended in 1mM HEPES containing 10% glycerol. Km^r colonies were selected on LB plates containing Km (250 µg/ml). To select for double crossover mutants, a second screening on LB plates containing Ap (300 µg /ml) was carried out. The inactivation of *cpnB* (Fig. 2), was confirmed by PCR. The resulting mutant HI-201 was found not to be able to grow on cyclopentanol or cyclopentanone as a sole carbon and energy source. This result indicated that *cpnB* is essential for the degradation of cyclopentanol and it appeared that there was only one copy of the *cpnB* gene in strain 9872.

As expected of a flavoprotein the amino acid sequence of CPMO contains motifs of FAD fingerprints similar to those found in flavoprotein hydroxylases.

Nucleotide sequence accession number

The DNA sequence of the 4,281-bp *Sph*I fragment has been submitted to DDBJ and assigned accession number AB022102. The release of this data awaits the inventors' authorization.

While the invention has been described in connection with specific embodiments thereof, it will be understood that it is capable of further modifications and this application is intended to cover any variations, uses, or adaptations of the invention following, in general, the principles of the invention and including such departures from the present disclosure as come within known or customary practice within the art to which the invention pertains and as may be applied to the essential features hereinbefore set forth, and as follows in the scope of the appended claims.

WHAT IS CLAIMED IS:

1. An isolated DNA encoding a cyclopentanone monooxygenase (CPMO), or an enzymatically active portion thereof, the isolated DNA being characterized by the ability to hybridize specifically with the complement of the DNA represented in SEQ ID NO:8 under stringent hybridization conditions.
2. An isolated DNA, wherein it codes for a cyclopentanone monooxygenase (CPMO), and contains:
 - (a) the nucleic acid sequence of SEQ ID NO:8;
 - (b) a sequence corresponding to said nucleic acid sequence in the scope of the degeneration of the genetic code; or
 - (c) a sequence hybridizing under stringent conditions with the sequence from (1) or (2), and still coding for cyclopentanone monooxygenase (CPMO).
3. An isolated DNA encoding a cyclopentanone monooxygenase (CPMO), or an enzymatically active portion thereof, said isolated DNA having SEQ ID NO:8.
4. An isolated DNA expression vector encoding an enzymatically active cyclopentanone monooxygenase (CPMO) comprising a DNA characterized by a sequence as set forth in SEQ ID NO:8, or a portion thereof, said portion encoding said CPMO, in expressible form.
5. A recombinant vector comprising the isolated DNA of any one of claims 1 to 3, wherein the isolated DNA encodes cyclopentanone monooxygenase.
6. The recombinant vector of claim 5, wherein the isolated DNA has a nucleic acid sequence of SEQ ID NO:8 or which, due to the degeneracy of the genetic code, is a functional equivalent thereof.
7. A recombinant vector, wherein it contains one or more copies of a recombinant DNA according to claim 1, 2 or 3.
8. A recombinant vector according to claim 4, 5, 6 or 7, wherein it is a prokaryotic vector.

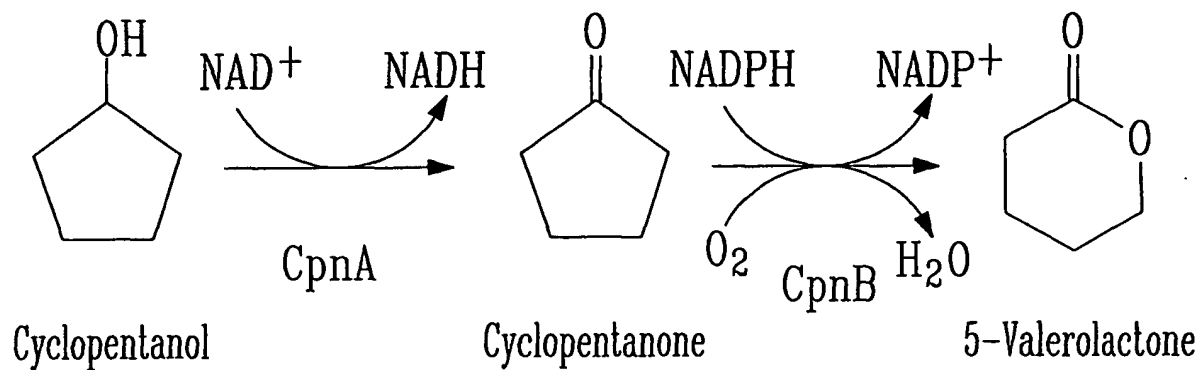
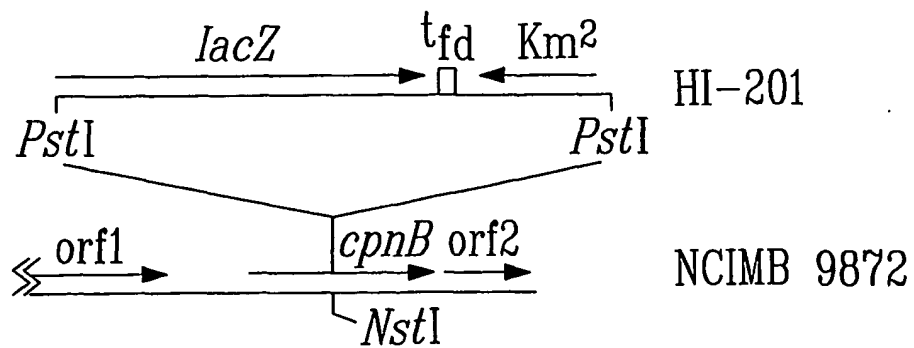
9. A recombinant vector according to claim 4, 5, 6 or 7, wherein it is a plasmid.
10. Biologically functional plasmid or viral DNA vector, which contains a DNA as defined in claim 1, 2 or 3.
11. A host cell comprising a recombinant vector of any one of claims 4 to 9.
12. A cell transformed with a heterologous DNA expression construct encoding an enzymatically active cyclopentanone monooxygenase (CPMO) comprising a DNA characterized by a sequence as set forth in SEQ ID NO:8, or a portion thereof, said portion encoding said CPMO, in expressible form.
13. A cell as defined in claim 12, which is a prokaryotic cell.
14. A cell as defined in claim 12, which is E. coli.
15. A purified cyclopentanone monooxygenase (CPMO) having :
 - (a) an amino acid sequence as set forth in SEQ ID NO:5;
 - (b) an amino acid sequence encoded by a nucleic acid sequence as set forth in SEQ ID NO:8; or
 - (c) an amino acid sequence encoded by a nucleic acid sequence hybridizing to a nucleic acid sequence complementary to the nucleic acid sequence of step b) above under stringent conditions, said amino acid sequence encoded in step c) having a same activity as the amino acid sequence in a).
16. A recombinant cyclopentanone monooxygenase (CPMO) having an enzymatic activity superior to the one naturally occurring.
17. The recombinant cyclopentanone monooxygenase (CPMO) of claim 16, wherein said CPMO is prepared from Comamonas sp. NCIMB 9872.
18. The recombinant cyclopentanone monooxygenase (CPMO) of claim 16, wherein said CPMO has a sequence as set forth in SEQ ID NO:5.

19. The recombinant cyclopentanone monooxygenase (CPMO) of claim 16, wherein said CPMO has an enzymatic activity twice superior to that of a CPMO from a native *Pseudomonas*.

20. A method for growing cells in vitro in presence of cyclopentanol or cyclopentanone as sole source of carbon, said method comprising the steps of:

- (a) transforming a cell with the expression construct of claim 4, 5, 6, 7, 8 or 9; and
- (b) growing the cell of step a) under suitable conditions in a medium containing cyclopentanol or cyclopentanone as a sole source of carbon.

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FIG. 1FIG. 2

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GxGxxG

CHMO 1:-----MSQKMDFDAIVIGGGFGGLYAVKKLRDELELKVQAFDKATDVAGT 45

STMO 1:MNGQHPRSVVTAPDATTGTTSDYDVVVVGAGIAGLYAIHRFRSQ-GLTVRAFEAASGVGGV 59

CPMO 1:MTTFTMTTEQLGMNNSVNDKLDVLLIGAGFTGLYQLYHLRKL-GYKVHLVDAGADIGGI 59

. * ...* *. **** * . . . *

CHMO 46:WYWNRYPGALTDTEHLYCYSWDKELLQSLEIKKKYVQGPDVRKYLQQVAEKHDLKKSQ 105

STMO 60:WYWNRYPGARCDVESIDYSYSFSPELEQEWNVSEKYATQPEILAYLEHVADRFDLRRDIR 119

CPMO 60:WHWNCYPGARVDTHCQIYQYSI-PELWQEFNWKEFPNWAQMREYFHFADKKLDLSKDIS 118

*. ** ***** * * ** * * * . . ** .

CHMO 106:FNTAVQSAHYNEADALWEVTTEYGDKYTARFLITALGLLSAPNLPNIKGINQFKGELHHT 165

STMO 120:FDTRVTSAVLDEEGLRWTVRTRDGRDEV SARFLVVAAGPLSNANTPAFDGLDRFTGDIVHT 179

CPMO 119:FNTRVQSAVFDEGTREWTVRSIGHQPIQARFVIANLGFGASPSTPNVDGIETFKGQWYHT 178

*. * * * * . * * * . . * * * . * . * * . * * . **

GxGxxG

CHMO 166:SRWPDD-VSFEGKRVGVIGTGSTGVQVITAVAPLAKHLTVFQRSAQYSVPIGNDPLSEED 224

STMO 180:ARWPHDGVDFGTGKRVGVIGTGSSGIQSIPIIAEQAEQLFVFQRSANY SIPAGNVPLDDAT 239

CPMO 179:ALWPQEGVNMAGKRVAIIGTGSSGVQVAQEALDAKQVTVYQRTPNLALPMHQQLSAED 238

. ** * . ***** . ***** . * . * * * . * * . . . * . *

CHMO 225:VKKIKDNYDKSLGWCMSALAFALNESTVPAMSVSAEERKAVFEKAWQTGGGFRFMFETF 284

STMO 240:RAEQKANYAERRRLSRESGGGSPHRPHPKSALEVSEEERRAVYEERWKLGG--VLFSKAF 297

CPMO 239:NLRMKPELPAAAFERRGKCFAGFD FDI AKNATELSAAERTEILEELWNAGG-FRYWLANF 297

* . . * . * * * . * . * * * . *

FILE 3A

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CHMO 285:GDIATNMEANIEAQNFIFKKGIAEIVKDPAlAQKLPQD----LYAKRPLCDSGYNTFNR 340

STMO 298.PDQLTDPAAANDTARAFWEEKIRAVVDDPAVAELLTPKDH--AIGAKRIVTDSGYYETYNR 355

CPMO 298:QDYLFDDKANDYVYEFWRDKVRARIKDPKVAEKLAPMKKPHPYGAKRPSLEQWYYEIFNQ 357

* . ** * * . ** * . * * * . ** . *

CHMO 341.DNVRLEDVKANPIVEITENGVKLENGDFVELDMLICATGFDAGNYVRMDIQKNGLAM 400

STMO 356:DNVELVDLRSTPIVGMDGTGIVT-TGAHYDLDMIVLATGFDAMTGSCLKLEIVGRGGRTL 414

CPMO 358:NNVTLVDPNETPVLRITEKGIVT-AEGEAEFDLIVFATGFDAGTGGITSIDFRNNQGSF 416

. ** * * . * . . * * . . * . . * . . . * . .

CHMO 401:KDYWKEGPSSYMGVTVNNYPNMFVLGPNP--FTNLPPSIESQVEWISDTIQYTVENNV 458

STMO 415:KETWAAGPRTYLGLGIDGFPNFFNLTPGSPSVLANMVLHSELHVDWVADAIAYLDARGA 474

CPMO 417:KDVWSDGIRTQLGVATAGFPNLLFGYGPQSPAGFCNGPSSAEYQGDLILQLMNYLRDNNI 476

* . * * . * . . ** . . * * * . * * *

CHMO 459:ESIEATKEAEEQWTQTCANIAEMTLFPAQSWIFGANIPGKKNTVYFYLGLKEYRTCAS 518

STMO 475:AGIEGTPEAVADWVEECRNRAEASLLNSANSWYLGANIPGRPRVFMPLGGFGVYREIIT 534

CPMO 477:SRIEAQSEAQEEWSKLIADFWDSSLFPRAKSWYQGSNIPGKKVESLNFPLGLPTYISKFN 536

** ** . * . . . * . * . ** * . * . . . *

CHMO 519:NCKNHAYEGFDIQLQRSDIKQPANA 543 SEQ ID NO:7

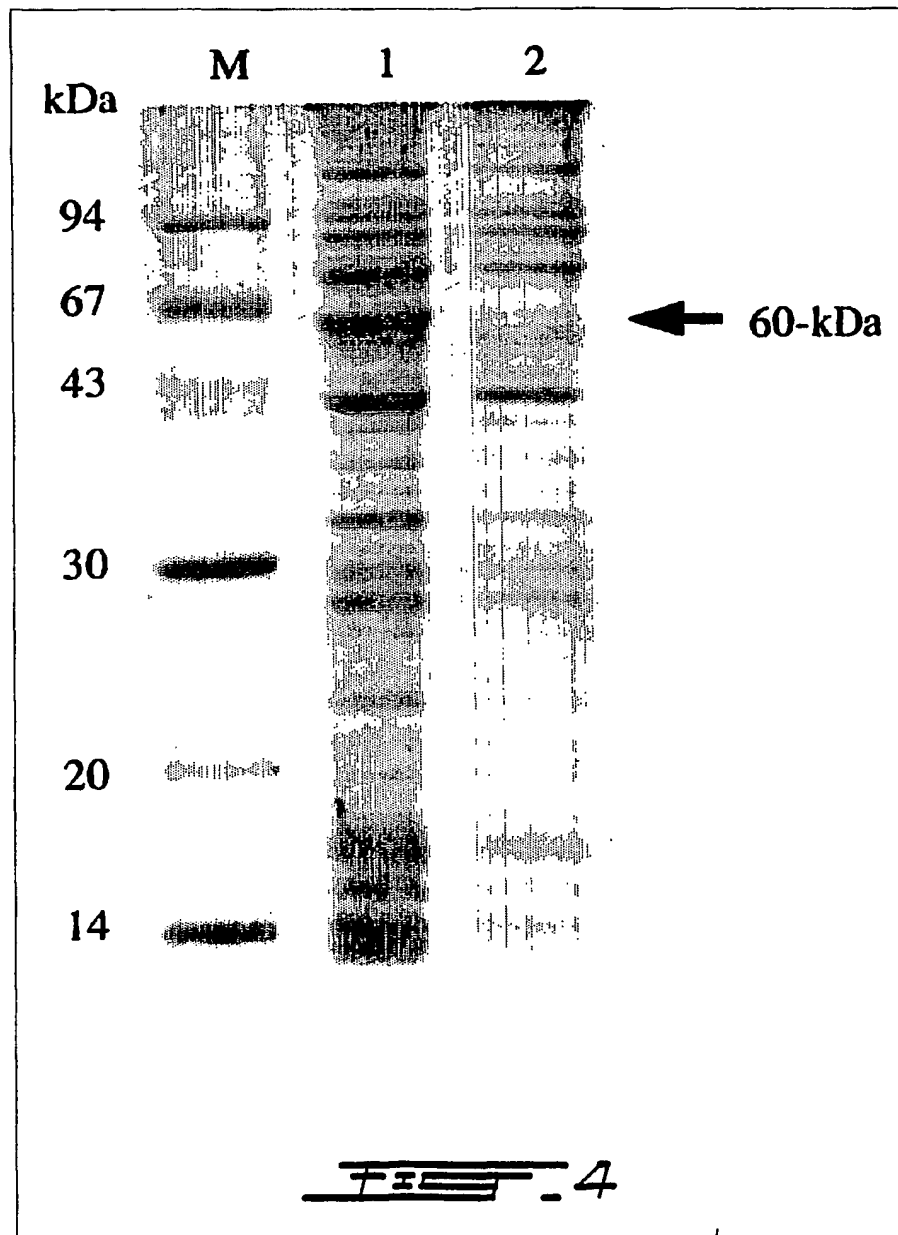
STMO 535:EVAESGYKGFAILEG----- 549 SEQ ID NO:6

CPMO 537:ESAEGYAGFSLAS----- 550 SEQ ID NO:5

. . * ** .

FE 3B

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	10	20	30	40	50	60
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	CGTACGCTATCGCCGTGCAACCACCCGCCCCTCGCGCCGGTGCTCGCGCAGAGAGCGAG					
61	CACCGACATCACATCGCCGGTCACCGGCGAGAACATTGGCAAGATCCTGCGCTTTTCGCC					
	GTGGCTGTAGTGTAGCGGCCAGTGGCCGCTCTTGTAACCGTTCTAGGACGCGAAAAGCGG					
121	GCTGAAGCCGCAGGGCTACGAGAGAATTTTTCAGTCCGGCCTGGCGGACTCGACCCAGCA					
	CGACTTCGGCGTCCCGATGCTCTCTTAAAAAGTCAGGCCGGACCGCCTGAGCTGGGTCTGT					
181	CACCCCCAGGATGGTGCCTTTTCGGACAGTTCCTGTCTGTGATCCAGAGACCATCAACAC					
	GTGGGGGTCTACCACGCAAAGCCTGTCAAGGACAGGACACTAGGTCTCTGGTAGTTGTG					
241	CCTGGAGACATTGGAGCGGATCGCGCGCGCCGACGTCAATGTCTTGCTGCATGGAGAGAC					
	GGACCTCTGTAACCTCGCCTAGCGCGCGGGCTGCAGTTACAGAACGACGTACCTCTCTG					
301	GGGCACGGGCAAAGAGCTGATAGCTCGCCACATTCATGTCGCGAGCCGCCGGCGGGATGC					
	CCCGTGCCCGTTTCTCGACTATCGAGCGGTGTAAGTACAGCGCTCGGCGGCCGCCCTACG					
361	GCCCTACCTCGCCATCAACTGCGGGGCAATCAGCTCGGAGTTGCTGGAGAGTACTTTTTT					
	CGGGATGGAGCGGTAGTTGACGCCCCGTTAGTCGAGCCTCAACGACCTCTCATGAAAAAA					
421	TGGCTATGTGCGCGGAGCATTCTCCGGGGCAGATCCCAAGGGCCGCGCCGGCTACTTTGA					
	ACCGATACACGCGCCTCGTAAGAGGCCCCGTCTAGGGTTCCCGGCGCGGCCGATGAACT					
481	ATCGGTGGGCGAAGGCACCTTGTTTCTGGACGAGATCGGCGAGCTGCCCTTGGCCATGCA					
	TAGCCACCCGCTTCCGTGGAACAAAGACCTGCTCTAGCCGCTCGACGGGAACCGGTACGT					
541	GGCGGCATTGTTGCGCGTGCTGGAAGACGGAAGCTTTCTGCGTGTGGGCTCATCGACCCC					
	CCGCCGTAACAACGCGCACGACCTTCTGCCTTCGAAAGACGCACACCCGAGTAGCTGGGG					
601	GCAGCGCGCCTCGTGCCGCATTATCGCGGCCACGCACCGCAATCTGGAGGAACATCGC					
	CGTCGCGCGGAGCACGGCGTAATAGCGCCGGTGCGTGGCGTTAGACCTCCTTGAGTAGCG					
661	ACAGGGCCTGTTTCGCCAGGATCTCTACTACCGTCTCAAGATCGTTCAGAAAAGGCTCAA					
	TGTCCCGGACAAAGCGGTCCTAGAGATGATGGCAGAGTTCTAGCAAGTCTTTTCCGAGTT					

FIG. 5A

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721 GCCGATACGGGAACGCACCTGTGACATCGCCTTGCTGGCAGAGCAATTCAAATCTGCACT
CGGCTATGCCCTTGCGTGGACACTGTAGCGGAACGACCGTCTCGTTAAGTTTAGACGTGA
781 GGCGCAAAAGCACACAGATCCCGAACGTGCAGATCCACCCGAGGCCATGGCAGTCATGGA
CCGCGTTTTTCGTGGTCTAGGGCTTGACGTCTAGGTGGGGCTCCGGTACCGTCAGTACCT
841 GCGCTACCAGTGGCCGGGCAATGCGCGCGAAATCCGCAATGTGATGGAAGCCGCCCTGAT
CGCGATGGTCACCGGCCCCGTACGCGCGCTTTAGGCGTTACACTACCTTCGGCGGGACTA
901 CTGCTCCGATGGTGAAATCACGCTCGCCAGCCTGCCCCGGAAGTATCTGAGAACTCCAC
GACGAGGCTACCACTTTAGTGCGAGCGGTCGGACGGGGGCCTTCATAGACTCTTGAGGTG
961 CTATCCTCTGCAAAGCCGGGTGGCCGAAGCGAATAGCGAGATGCCTCCCGTCTCCGGCAA
GATAGGAGACGTTTCGGCCCACCGGCTTCGCTTATCGCTCTACGGAGGGCAGAGGCCGTT
1021 CGACTATGAACGGCAACTCATCGTGGGTTTGCTGCGCAAGTACCGAAAGGTCAATCATGT
GCTGATACTTGCCGTTGAGTAGCACCCAAACGACGCGTTCATGGCTTTCCAGTTAGTACA
1081 GGCGAGGGCCCTGGGCCTCGCGCGCTCCACGCTCTATCGGAAATTCGCAGACTTGGGCAT
CCGCTCCCGGGACCCGGAGCGCGGAGGTGCGAGATAGCCTTTAAGCGTCTGAACCCGTA
1141 TGACCAACGCGAGTATGTCAATGATCTGTCCGATTAACACTGCTGCGCCCTGCGCTTGCG
ACTGGTTGCGCTCATACAGTTACTAGACAGGCTAATTGTGACGACGCGGGACGCGAACGC
1201 GCAACGCAGTATCGTCGGAACGCGGGCAACGCTAAAGGCAGCCGCCGTGCGCATCGGTTG
CGTTGCGTCATAGCAGCCTTGCGCCCGTTGCGATTTCCGTGCGCGGCAGCCGTAGCCAAC
1261 CCGCCCGGTGTTGTTGTGTCGGCTGAGACCGCCTGCAGCACCAGCCCCTCCATTCTCGTG
GGCGGGCCACAACAAACAGCCGACTCTGGCGGACGTCGTGGTCGGGGAGGTAAGGAGCAC
1321 TCTAGATCTCCTTGCAACGCTGTACAGCGTGCTTTTTTCGTGGCGCGGGACTTCTCGCG
AGATCTAGAGGAACGTTGCGACAGTGTCGCACGAAAAAGCACCGCGCCCTGAAGAGCGC
1381 ATGAGGGCCTGCAAGGGCGGCGCATCAAACAAGCGCCCTGTCCTCAGGACGTTTGCGCC
TACTCCCGGACGTTCCCGCCGCGTAGTTTTGTTGCGGGACAGGAGTCCTGCAAACGCGG
1441 GTCGCTGGCTATGGGCAGTCTGCATGATCCGCTTGGGCTTGCCCACCATGCGCGCGACTG

A stylized signature or logo consisting of a horizontal line with a series of vertical strokes underneath, followed by the letters "SB".

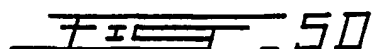
7/10

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1501 CGCTCTTTCTGTGGGCGCATCGTCATCCCGACCTTCCTGAAATCCATTTCCACAGGGC
GCGAGAAAGGACAACCCGCGTAGCAGTAGGGCTGGAAGGACTTTAGGTAAAGGTGTCCCG
1561 CGGCCTGAGCCAGACAGCGATCGGCAGGCTCTCGCATCTCGGACAGTGGTGCGCTACACC
GCCGGACTCGGTCTGTGCTAGCCGTCCGAGAGCGTAGAGCCTGTCACCACGCGATGTGG
1621 ACAGGACACAAGTGTCCGGCGCGTGGCACGCAGCGGCCCTTTGCTGCGCACGATGCGTGCT
TGTCTGTGTTTACAGGCCGCGCACCGTGCCTGCGCCGAAACGACGCGTGCTACGCACGA
1681 TTTGGCACATGGAAACCCCTCGGTTTTTCCGAACCAGAGACCTGGCACAAGTCTTGATAAC
AAACCGTGACCTTTGGGAGCCAAAAGGCTTGGTCTCTGGACCGTGTTTCAAGTATTTG
1741 TCCTGAGTGACTGATGCAACGCATCTCAGTCCGGGAGCGTGGACGACACGCCCTGAAC
AGGACTCACTGACTACGTTGCGTAGAGTCAGGCCCTCGCACCTGCTGTGCGGGGACTTGA
1801 TCAATTATTTAGGAGACCCATATGACCACCATGACCACCATGACCACCGAACAACCTCGGC
AGTTAATAAATCCTCTGGGTATACTGGTGGTACTGGTGGTACTGGTGGCTTGTGAGCCG
1861 ATGAACAACCTCTGTCAATGACAAGCTTGACGTTTTGCTCATCGGCGCCGGCTTCACCGGT
TACTTGTTGAGACAGTTACTGTTGAACTGCAAAACGAGTAGCCGCGGCCGAAGTGGCCA
1921 CTCTACCAGCTCTATCACCTGCGCAAGCTGGGCTACAAGGTTCTCTCGTCGACGCCGGT
GAGATGGTCGAGATAGTGGACGCGTTTCGACCCGATGTTCCAAGTAGAGCAGCTGCGGCCA
1981 GCCGATATTGGCGGGATCTGGCATTGGAACCTGCTACCCCGAGCGCGTGTGGATACCCAC
CGGCTATAACCGCCCTAGACCGTAACCTTGACGATGGGGCCTCGCGCACACCTATGGGTG
2041 TGCCAGATCTACCAGTACTCCATTCCAGAGTTGTGGCAGGAGTTCAACTGGAAAGAGCTG
ACGGTCTAGATGGTCATGAGGTAAGGTCTCAACACCGTCCTCAAGTTGACCTTTCTCGAC
2101 TTCCCTAACTGGGCGCAAATGCGCGAGTATTTCCATTTTGCCGACAAGAAGCTCGACCTG
AAGGGATTGACCCGCGTTTACGCGCTCATAAAGGTAAAACGGCTGTTCTTCGAGCTGGAC
2161 AGCAAGGACATCAGCTTCAACACCCGTGTGCAGTCGGCCGTCTTTGACGAAGGCACACGC
TCGTTCCCTGTAGTCGAAGTTGTGGGCACACGTCAGCCGGCAGAACTGCTTCCGTGTGCG

FIG. 5C

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2221 GAATGGACGGTACGCTCGATCGGACACCAGCCGATCCAGGCCAGGTTCGTTCATCGCCAAC
CTTACCTGCCATGCGAGCTAGCCTGTGGTCGGCTAGGTCCGGTCCAAGCAGTAGCGGTTG
2281 CTTGGCTTCGGTGCCAGCCCCAGCACGCCCAATGTCGATGGCATCGAGACATTCAAAGGC
GAACCGAAGCCACGGTCGGGGTTCGTGCGGGTTACAGCTACCGTAGCTCTGTAAGTTTCCG
2341 CAGTGGTATCACACTGCTCTGTGGCCCCAGGAAGGCGTGAACATGGCCGGCAAGCGCGTG
GTCACCATAGTGTGACGAGACACCGGGTCTTCCGCACTTGTACCGGCCGTTTCGCGCAC
2401 GCCATCATTGGCACCGGCTCCAGCGGGTCCAGGTCGCCCAGGAGGCTGCCCTTGATGCG
CGGTAGTAACCGTGGCCGAGGTCGCCCCAGGTCCAGCGGGTCTCCGACGGGAACCTACGC
2461 AAACAGGTGACGGTGTACCAGCGCACCCCCAACCTGGCCTTGCCCATGCATCAGAAGCAG
TTTGTCCACTGCCACATGGTCGCGTGGGGGTTGGACCGGAACGGGTACGTAGTCTTCGTC
2521 CTCAGCGCCGAGGACAATCTGCGCATGAAGCCCGAGCTTCCGCGAGCGTTCGAGAGACGC
GAGTCGCGGCTCCTGTTAGACGCGTACTTCGGGCTCGAAGGGCGTCGCAAGCTCTCTGCG
2581 GGCAAGTGCTTCGCCGGCTTCGACTTCGACTTCATCGCCAAGAACGCGACCGAGCTGTCC
CCGTTACGAAGCGGCCGAAGCTGAAGCTGAAGTAGCGGTTCTTGCGCTGGCTCGACAGG
2641 GCTGCGGAGCGCACAGAGATCTTGAAGAGCTGTGGAACGCCGGCGGCTTCGGCTACTGG
CGACGCCTCGCGTGCTCTAGAACCTTCTCGACACCTTGCGGCCGCCGAAGGCGATGACC
2701 CTGGCCAATTTCCAAGACTATCTGTTTCGATGACAAGGCCAACGATTACGTCTACGAGTTC
GACCGGTTAAAGGTCTGATAGACAAGCTACTGTTCCGGTTGCTAATGCAGATGCTCAAG
2761 TGGCGCGACAAGGTCCGCGCCCGCATCAAGGATCCGAAAGTTGCCGAGAAGCTCGCCCCC
ACCGCGCTGTTCCAGGCGCGGGCGTAGTTCCTAGGCTTTCAACGGCTCTTCGAGCGGGGG
2821 ATGAAGAAGCCGCATCCTTACGGAGCCAAGCGCCCTTCGCTGGAGCAGTGGTACTACGAG
TACTTCTTCGGCGTAGGAATGCCTCGGTTTCGCGGAAGCGACCTCGTCACCATGATGCTC
2881 ATCTTCAATCAGAACACGTCACGCTGGTGGATGTCAACGAAACACCGGTGCTTCGCATC
TAGAAGTTAGTCTTGTTGCAGTGCAGCACCTACAGTTGCTTTGTGGCCACGAAGCGTAG
2941 ACCGAGAAAGGCATCGTGACCGCTGAGGGTGAAGCCGAATTCGACCTGATCGTGTTTCGCG

The logo consists of the word "FIS" in a stylized, bold, sans-serif font, followed by the number "50". The "FIS" is underlined, and the "50" is also underlined. The entire logo is rendered in a dark, possibly black or dark grey, color.

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TGGCTCTTTCCGTAGCACTGGCGACTCCCACTTCGGCTTAAGCTGGACTAGCACAAAGCGC
3001 ACCGGCTTCGACGCAGTGACCGGGGGACTCACCAGCATCGACTTCCGCAACAACCAGGGC
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3121 GGTTCCTCCCACTTGCTCTTTGGCTACGGACCTCAATCGCCTGCGGGCTTCTGCAACGGT
CCAAAAGGGTTGAACGAGAAACCGATGCCTGGAGTTAGCGGACGCCCCAAGACGTTGCCA
3181 CCGTCGAGCGCCGAATACCAGGGCGATCTGCTGATCCAGCTGATGAACTACCTACGCGAC
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3241 AACAACATCTCGCGCATCGAAGCCCAGTCCGAGGCACAGGAAGAATGGAGCAAGCTGATC
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3301 GCAGACTTCTGGGACAGCTCGCTGTTCCCCCGCGCAAAGTCCTGGTACCAAGGATCCAAC
CGTCTGAAGACCCGTGTCGAGCGACAAGGGGGCGCGTTTCAGGACCATGGTTCCTAGGTTG
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3541 CTGGCGGTGCCATGGGAATGGGCCTGACCCATTGCACCTTGCTGGCCCCGCGAGGGGGCCA
GACCGCCACGGTACCCTTACCCGGACTGGGTAACGTGGAACGACCGGGCGCTCCCCCGGT
3601 CCGTTTACCTCAGCGACATGAATGAGGAACTGGGCCACCAAGCCGTAGCGGAGATCCGTC
GGCAAATGGAGTCGCTGTACTTACTCCTTGACCCGGTGGTTTCGGCATCGCTCTAGGCAG
3661 GGCAAGGTGGCAAGGCCCACTTCCTGCACCTGGACGTGACCAACGAGAATCACTGGACAG
CCGTTCCACCGTTCGGGTGAAGGACGTGGACCTGCACTGGTTGCTCTTAGTGACCTGTC

A stylized signature or logo consisting of a horizontal line with a small vertical tick on the left, followed by the letters "SE" in a bold, sans-serif font.

10/10

3721 GTGCCGTGGACACCATCCTCGCCGAGTCCGACCGGCTGGACGCGCTGGTCAACAACGCCG
CACGGCACCTGTGGTAGGAGCGGCTCAGGCTGGCCGACCTGCGCGACCAGTTGTTGCGGC
3781 GCATTTTGACCCTCAAACCCGTTCAAGACACCAGCAACGAAGAATGGGATCGCATCTTTG
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3841 AAATCAATGTCCGCAGTGTGTTCTTGGGCACGCGAGCAGTGATCGAGCCCATGCGTAAAG
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4201 AGGCCTCGTTCGTGCATGGCTCTGAACTCGTCGTCGACGGAGGGTATACGGCGAACTGAG
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SEQUENCE LISTING

<110> NATIONAL RESEARCH COUNCIL OF CANADA

IWAKI, Hiroaki

HASEGAWA, Yoshie

LAU, Peter C.K.

<120> Cloning, Sequencing and Expression of a
Comamonas Cyclopentanone 1,2-Monooxygenase-Encoding Gene in
Escherichia coli

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<151> 2000-07-18

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35 40 45
Leu Val Asp Ala Gly Ala Asp Ile Gly Gly Ile Trp His Trp Asn Cys
50 55 60
Tyr Pro Gly Ala Arg Val Asp Thr His Cys Gln Ile Tyr Gln Tyr Ser
65 70 75 80
Ile Pro Glu Leu Trp Gln Glu Phe Asn Trp Lys Glu Leu Phe Pro Asn
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100 105 110
Leu Ser Lys Asp Ile Ser Phe Asn Thr Arg Val Gln Ser Ala Val Phe
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130 135 140
Ile Gln Ala Arg Phe Val Ile Ala Asn Leu Gly Phe Gly Ala Ser Pro
145 150 155 160
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165 170 175
His Thr Ala Leu Trp Pro Gln Glu Gly Val Asn Met Ala Gly Lys Arg
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210 215 220

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 Lys Ala Asn Asp Tyr Val Tyr Glu Phe Trp Arg Asp Lys Val Arg Ala
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 Arg Ile Lys Asp Pro Lys Val Ala Glu Lys Leu Ala Pro Met Lys Lys
 325 330 335
 Pro His Pro Tyr Gly Ala Lys Arg Pro Ser Leu Glu Gln Trp Tyr Tyr
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 Glu Ile Phe Asn Gln Asn Asn Val Thr Leu Val Asp Val Asn Glu Thr
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 Pro Val Leu Arg Ile Thr Glu Lys Gly Ile Val Thr Ala Glu Gly Glu
 370 375 380
 Ala Glu Phe Asp Leu Ile Val Phe Ala Thr Gly Phe Asp Ala Val Thr
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 Gly Gly Leu Thr Ser Ile Asp Phe Arg Asn Asn Gln Gly Gln Ser Phe
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 Lys Asp Val Trp Ser Asp Gly Ile Arg Thr Gln Leu Gly Val Ala Thr
 420 425 430
 Ala Gly Phe Pro Asn Leu Leu Phe Gly Tyr Gly Pro Gln Ser Pro Ala
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 450 455 460
 Ile Gln Leu Met Asn Tyr Leu Arg Asp Asn Asn Ile Ser Arg Ile Glu
 465 470 475 480
 Ala Gln Ser Glu Ala Gln Glu Glu Trp Ser Lys Leu Ile Ala Asp Phe
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 Trp Asp Ser Ser Leu Phe Pro Arg Ala Lys Ser Trp Tyr Gln Gly Ser
 500 505 510
 Asn Ile Pro Gly Lys Lys Val Glu Ser Leu Asn Phe Pro Leu Gly Leu
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 Ala Phe Glu Ala Ala Ser Gly Val Gly Gly Val Trp Tyr Trp Asn Arg
 50 55 60
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